

Communication

A New Strategy to Induce D-Turns: Peptides Composed of Alternating D-Aminoxy Acids and D-Amino Acids

Dan Yang, Wei Li, Jin Qu, Shi-Wei Luo, and Yun-Dong Wu

J. Am. Chem. Soc., **2003**, 125 (43), 13018-13019• DOI: 10.1021/ja036136p • Publication Date (Web): 02 October 2003 Downloaded from http://pubs.acs.org on March 30, 2009



More About This Article

Additional resources and features associated with this article are available within the HTML version:

- Supporting Information
- Links to the 8 articles that cite this article, as of the time of this article download
- Access to high resolution figures
- Links to articles and content related to this article
- Copyright permission to reproduce figures and/or text from this article

View the Full Text HTML





Published on Web 10/02/2003

A New Strategy to Induce γ -Turns: Peptides Composed of Alternating α -Aminoxy Acids and α -Amino Acids

Dan Yang,*,†,§ Wei Li,† Jin Qu,† Shi-Wei Luo,‡ and Yun-Dong Wu*,‡

Department of Chemistry, The University of Hong Kong, Pokfulam Road, Hong Kong, China, Department of Chemistry, Fudan University, Shanghai, China, and Department of Chemistry, The Hong Kong University of Science and Technology, Clear Water Bay, Kowloon, Hong Kong, China

Received May 14, 2003; E-mail: yangdan@hku.hk; chydwu@ust.hk

As one type of reversed turn secondary structures of proteins, γ -turns are formed by a 3 \rightarrow 1 hydrogen bond between the CO group of amino acid residue *i* and the NH group of amino acid residue *i* + 2.¹ While γ -turns are less frequently observed in proteins than β -turns,^{1c} they have been shown to play important roles in biological recognition.² For example, the γ -turn present in RGD (Arg-Gly-Asp) sequence of vitronectin was found to contribute to the specific recognition by integrin receptor $\alpha_{V}\beta_{3}$.³ However, it has been challenging to investigate the roles of γ -turns in protein—peptide recognition because they seldom exist in short, linear peptides. Most of the γ -turn mimics constrain the peptide conformation through ring formation.^{4,5} Here we report that the γ -turns can be initiated by the following N—O turns, an eight-membered-ring intramolecular hydrogen bond induced by an α -aminoxy acid.⁶



Peptides 1-5 composed of alternating L-aminoxy value and D-alanine were prepared according to a convergent synthetic scheme. In 2 and 3, an α -amino acid was placed before or after an α -aminoxy acid, whereas in 4 and 5, each α -amino acid was sandwiched by two adjacent α -aminoxy acids.

Chemical shifts of amide NHs of 1-5 taken in CD₂Cl₂ at very low concentration (1-2 mM) are summarized in Table 1. For the regular amides, only the NH_a of dipeptide **2** and NH_c of dipeptide **3** fell in the range of non-hydrogen-bonded amide protons. In contrast, those regular amide NHs immediately after an α -aminoxy acid residue in 1-5 were downfield shifted by about 2 ppm, suggesting that they form strong intramolecular hydrogen bonds, most likely the N-O turns. For the *N*-terminal aminoxy amide NHs of peptides **1** and **3**-**5**, they appeared in the range of 8.36-8.48 ppm, assigned to be non-hydrogen-bonded NHs. The unusually downfield chemical shifts of the other aminoxy amide protons (δ 9.63-9.83 ppm) suggested the formation of seven-membered-ring intramolecular hydrogen bonds (the γ -turn) between aminoxy amide NH_i and C=O_{i-2}. The higher acidity of aminoxy amide protons may contribute to the formation of the observed γ -turns.

¹H NMR studies of peptides **1–5** were also carried out at 10 mM in CD₃OH at -30 °C (Table 1). For each peptide, the hydrogen-bonding patterns in CD₃OH were found to be similar to those in nonpolar solvent CD₂Cl₂. The chemical shifts of NH_a of **1**

Table 1.	Chemical Shifts	of Amide	NHs of	1-5 in CD2	Cl ₂ at	25
C at Lov	v Concentration	(1-2 mM)	, and in	CD ₃ OH (10	mM)	at
−30 °C ^a						

	δHa	δH_b	δH_{c}	δH_d	δH_{e}	δH _r
	CD ₂ Cl ₂ CD ₃ OH	CD2Cl2 CD3OH	CD2Cl2 CD3OH	CD2Cl2 CD3OH	CD ₂ Cl ₂ CD ₃ OH	CD2Cl2 CD3OH
1	<u>8.36</u> <u>11.46</u>	8.15 8.94				
2	5.91 8.01	<u>9.63</u> <u>11.89</u>	7.94 8.90			
3	8.46 11.46	8.60 9.01	6.37 8.55			
4	<u>8.45</u> <u>11.42</u>	8.82 9.11	<u>9.66 11.93</u>	8.07 8.86		
5	<u>8.48</u> <u>11.44</u>	8.85 9.31	<u>9.83</u> <u>11.92</u>	8.45 8.93	<u>9.74 11.89</u>	8.02 8.85

^a Aminoxy amide NHs are underlined.



Figure 1. Summary of NOEs observed of compounds 4 and 5 at 25 °C (5 mM in CD₂Cl₂; s, strong NOE; m, medium NOE; w, weak NOE).

and 3-5 appeared at about 11.45 ppm, more downfield than in CD₂Cl₂ due to the formation of intermolecular hydrogen bond to CD₃OH, whereas the peaks of other aminoxy amide protons were downfield shifted to 11.9 ppm. Furthermore, after solvent suppression, the peaks of non-hydrogen-bonded aminoxy amide NH_a disappeared due to the fast exchange with solvent, while the signals of hydrogen-bonded aminoxy amides still remained but became weaker. As to the regular amide NHs, the chemical shifts of NH_a of **2** and NH_c of **3** were rather upfield, assigned to be free amides. Other regular amide NHs were all downfield shifted by more than 0.3 ppm compared to the free amide NH_c of **3**. These data suggested that the alternating N–O turns and γ -turns still exist in methanol.

NOESY results of oligomers 4 and 5 in CD₂Cl₂ are summarized in Figure 1. For each α -D-alanine residue, similar intensities of NOE between $C_{\alpha}H_i$ and NH_i and those between $C_{\alpha}H_i$ and NH_{i+1} suggested that the α -proton takes an axial position in the γ -turn, resulting in an inverse γ -turn conformation.^{1c,7} As to the aminoxy acid residues, according to our previous theoretical calculation, C_{α} - C_{β} bond is anti to the N–O bond in the most stable N–O turn conformation; thus, stronger NOE between NH_i and $C_{\alpha}H_i$ than that between NH_{i+1} and $C_{\alpha}H_i$ is expected.^{6c,d} This was indeed observed for the first and the last α -L-aminoxy acid residues of 4 and 5. Interestingly, another conformer of the left-handed N–O turn was observed in the middle α -L-aminoxy acid residue of pentapeptide 5, in which the NOE between H_c and H_3 was of similar intensity to that of the NOE between H₃ and H_d. This NOE pattern agreed with the second lowest-energy N-O turn conformation revealed by theoretical calculations.^{6d} In this conformer, the $O-C_{\alpha}$ bond was rotated slightly with the C_{α} - C_{β} bond gauche to the N-O bond. This conformational change probably rendered the two α -methyl

[†] The University of Hong Kong.

[§] Fudan University.

[‡] The Hong Kong University of Science and Technology.



Figure 2. Calculated, most stable conformation of compound **6** in CH₂Cl₂. Interatomic distances are in Å.



Figure 3. CD spectra of compounds 1-5 at 25 °C: (a) 0.4 mM in 2,2,2-trifluoroethanol; (b) 4.0 mM in methanol.

groups of the α -D-alanine residues parallel to each other so as to alleviate their steric repulsions.

Theoretical calculations have been carried out on several di-, tri-, and tetra-peptide models (see SI for detailes). These peptides have a stong tendency to form alternating N–O turn and γ -turn. Figure 2 shows the most stable conformation of model compound **6** in CH₂Cl₂. In the alanine residue, the α -methyl group took an equatorial position, and the dihedral angles (φ, ψ) for the γ -turn are (83.8, -88.1), characteristic of an inverse γ -turn.^{1c,7} For the aminoxy acid residues, the C $_{\alpha}$ –C $_{\beta}$ bond is anti to the N–O bond. The calculated short O- - -H distances and large O- - -H–N angles indicate the formation of strong hydrogen bonds. The calculated interatomic H/H distances agree well with the observed NOE pattern for tripeptide **4**. Overall, the three turns form an extended helix structure.

CD spectra of compounds 1-5 taken at room temperature in 2,2,2-trifluoroethanol are shown in Figure 3a. Because α -aminoxy acid backbone is predisposed to the N-O turn formation and the energy cost to initiate the secondary structure is lower than that of an α -amino acid, α -aminoxy acid residues are expected to make a major contribution to the CD absorption. Thus, the CD signals were normalized for the concentration and the number of backbone N-O turns of each compound. Peptide 3 showed a different CD curve compared with others, possibly because the γ -turn between NH_c and C=O of aminoxy acid of 3 was not formed as revealed by the above ¹H NMR studies. The CD curves of oligomers 4 and 5 were almost superimposable to each other with a maximum at 227 nm, a minimum at 195 nm, and a zero crossing at 211 nm, suggesting that oligomers 4 and 5 adopt the same type of secondary structure, a novel mixed 7-8 helix. CD curves of peptides 1-5 in methanol (Figure 3b) showed patterns similar to those in 2,2,2-trifluoroethanol, suggesting the secondary structures remain unchanged in methanol.

In summary, the conformational studies suggested that oligomers **4** and **5** form N–O turns and γ -turns simultaneously in solution, even in protic solvent methanol. ¹H NMR studies of peptides **2** and **3** implied that the γ -turn could only be initiated by the following

N–O turn,⁸ which means that this hydrogen bond must involve an acidic aminoxy amide NH. Therefore, we have developed a new strategy to induce a γ -turn at specific sites of short peptides by putting an α -aminoxy acid immediately after the particular α -amino acid of interest.

Acknowledgment. This work was supported by The University of Hong Kong, Hong Kong University of Science and Technology, and Hong Kong Research Grants Council. Croucher Foundation is acknowledged for Senior Research Fellowships (to D.Y. and Y.-D.W.). D.Y. thanks Bristol-Myers-Squibb for the Unrestricted Grants in Synthetic Organic Chemistry. We thank Professor M.-J. Zhang for advice on NMR studies in methanol.

Supporting Information Available: Synthetic scheme and characterization data of compounds 1-5; ¹H NMR studies of 4 and 5; ¹H NMR spectra of 1-5; 2D NOESY spectra of 4 and 5; theoretical calculations (PDF). This material is available free of charge via the Internet at http://pubs.acs.org.

References

- (a) Némethy G.; Printz, M. P. *Macromoleculers* **1972**, *5*, 755–758. (b) Toniolo, C. *Crit. Rev. Biochem.* **1980**, *9*, 1–44. (c) Rose, G. D.; Gierasch, L. M.; Smith, J. A. Adv. Protein Chem. **1985**, *37*, 1–109.
- (2) (a) Milon, A.; Miyazawa, T.; Higashijima, T. Biochemistry 1990, 29, 65–75. (b) Coles, M.; Sowemimo, V.; Scanlon, D.; Murro, S. L. A.; Craik, D. J. J. Med. Chem. 1993, 36, 2658–2665. (c) Grübler, G.; Echner, H.; Voelter, W.; Folkers, G.; Krug, M.; Siemion, I. Z. Pol. J. Chem. 1992, 66, 1269–1276. (d) Pędyczak, A.; Trojnar, J.; Siemion, I. Z. Pol. J. Chem. 1994, 68, 2181–2189. (e) Odell, B.; Hammond, S. J.; Osborne, R.; Goosey, M. W. J. Comput-Aided Mol. Des. 1996, 10, 89–99. (f) Carver, J. A.; Esposito, G.; Viglino, P.; Fogolari, F.; Guichard, G.; Briand, J.-P.; Van Regenmortel, M. H. V.; Brown, F.; Mascagni, P. Biopolymers 1997, 41, 569–590. (g) Porcelli, M.; Casu, M.; Lai, A.; Saba, G.; Pinori, M.; Cappelletti, S.; Mascagni, P. Biopolymers 1999, 50, 211–219. (h) Vogen, S. M.; Prakash, O.; Kirnarsky, L.; Sanderson, S. D.; Sherman, S. A. J. Pept. Res. 1999, 54, 74–84. (i) Andrianov, A. M.; Sokolov, Y. A. J. Biomol. Struct. Dyn. 2003, 20, 603–614.
- (3) (a) Aumailley, M.; Gurrath, M.; Müller, G.; Calvete, J.; Timpl, R.; Kessler, H. *FEBS Lett.* **1991**, 291, 50-54. (b) Haubner, R.; Gratias, R.; Diefenbach, B.; Goodman, S. L.; Jonczyk, A.; Kessler, H. J. Am. Chem. Soc. **1996**, *118*, 7461-7472. (c) Haubner, R.; Schmitt, W.; Hölzemann, G.; Goodman, S. L.; Jonczyk, A.; Kessler, H. J. Am. Chem. Soc. **1996**, *118*, 7881-7891. (d) Koppitz, M.; Huenges, M.; Gratias, R.; Kessler, H.; Goodman, S. L.; Jonczyk, A. Helv. Chim. Acta **1997**, *80*, 1280-1300. (e) Wermuth, J.; Goodman, S. L.; Jonczyk, A.; Helv. Chim. Acta **1997**, *80*, 1280-1300. (e) Wermuth, J.; Goodman, S. L.; Jonczyk, A.; Kessler, H. J. Am. Chem. Soc. **1997**, *119*, 1328-1335. (f) Burgess, K.; Lim, D.; Mousa, S. A. J. Med. Chem. **1996**, *39*, 4520-4526. (g) Schumann, F.; Müller, A.; Koksch, M.; Müller, G.; Sewald, N. J. Am. Chem. Soc. **2000**, *122*, 12009-12010.
- (4) For heterocyclic γ-turn mimics, see: (a) Etzkorn, F. A.; Travins, J. M.; Hart, S. A. Adv. Amino Acid Mimetics Peptidomimetics 1999, 2, 125– 163. (b) Alkorta, I.; Suarez, M. L.; Herranz, R.; González-Muñiz, R.; García-López, M. T. J. Mol. Model. 1996, 2, 16–25. (c) Callahani, J. F.; Bean, J. W.; Burgess, J. L.; Eggleston, D. S.; Hwang, S. M.; Kopple, K. D.; Koster, P. F.; Nichols, A.; Peishoff, C. E.; Samanen, J. M.; Vasko J. A.; Wong A.; Huffman, W. F. J. Med. Chem. 1992, 35, 3970–3972. (d) Newlander, K. A.; Callahan, J. F.; Moore, M. L.; Tomaszek, T. A., Jr.; Huffman, W. F. J. Med. Chem. 1993, 36, 2321–2331. (e) Callahan, J. F.; Newlander, K. A.; Burgess, J. L.; Eggleston, D. S.; Nichols, A.; Wong, A.; Huffman, W. F. Tetrahedron 1993, 49, 3479–3488.
- (5) For γ-turn induced by 2,3-methanoamino acids, see: (a) Burgess, K.; Ho, K.-K.; Petitit, B. M. J. Am. Chem. Soc. **1994**, *116*, 799–800. (b) Burgess, K.; Ke, C.-Y. J. Org. Chem. **1996**, *61*, 8627–8631. (c) Moye-Sherman, D.; Jin, S.; Li, S.; Welch, M. B.; Reibenspies, J.; Burgess, K. Chem.– Eur. J. **1999**, *5*, 2730–2739.
- (6) (a) Yang, D.; Ng, F.-F.; Li, Z.-J.; Wu, Y.-D.; Chan, W.-K.; Wang, D.-P. J. Am. Chem. Soc. 1996, 118, 9794–9795. (b) Yang, D.; Qu, J.; Li, B.; Ng, F.-F.; Wang, X.-C.; Cheung, K.-K.; Wang, D.-P.; Wu, Y.-D. J. Am. Chem. Soc. 1999, 121, 589–590. (c) Yang, D.; Li, B.; Ng, F.-F.; Yan, Y.-L.; Qu, J.; Wu, Y.-D. J. Org. Chem. 2001, 66, 7303–7312. (d) Wu, Y.-D.; Wang, D.-P.; Chan, K. W. K.; Yang, D. J. Am. Chem. Soc. 1999, 121, 11189–11196.
- (7) The inverse γ-turn is identified by an equatorial i + 1 side chain orientation while the classical γ-turn contains an axial i + 1 side chain. For p-amino acids, the inverse γ-turn with (φ,ψ) values generally appear in the range (70 to 95, -75 to -45).
- (8) For the corresponding L,L-isomer of 2, a γ-turn was also found. Unpublished results.

JA036136P